## Phosphorylation of Glucose with cyclo-Triphosphate

Mitsutomo TSUHAKO,\* Chiyoko SUEYOSHI, Yoshinobu BABA,† Tohoru MIYAJIMA,††
Shigeru OHASHI,†† Hiroyuki NARIAI,††† and Itaru MOTOOKA†††
Kobe Women's College of Pharmacy, Kitamachi, Motoyama, Higashinada-ku, Kobe
658

- † Department of Chemistry, Faculty of Education, Oita University, Dannoharu,
  Oita 870-11
- ††Department of Chemistry, Faculty of Science, Kyushu University 33, Hakozaki, Higashi-ku, Fukuoka 812
- †††Department of Chemisrty, Faculty of General Education, Kobe University,
  Tsurukabuto, Nada-ku 657

&-D-glucose reacted with cyclo-triphosphate ( $P_{3m}$ ) to give tri-, di-, and monophosphate derivatives. The main product, glucose tri-phosphate was a mixture of glucose 1- and 2-triphosphates (maximum yield = about 47%). Glucose was most effectively phosphorylated in alkaline solutions (pH 10—12) at room temperature.

Much attention has recently been focused upon cyclo-triphosphate  $(P_{3m})$  as a phosphorylating agent of organic compounds and a condensing agent of amino acids.  $P_{3m}$  readily reacts with ammonia,  $^{1,2}$ ) amines,  $^{3,4}$ ) amino acids,  $^{5-7}$ ) alcohols,  $^{8}$ ) phenols,  $^{9}$ ) and nucleosides  $^{10-15}$ ) to give phosphate derivatives. The present authors established a novel synthetic method of glucose triphosphate (glucose 1- and 2-triphosphates) by the reaction of  $\chi$ -D-glucose with  $P_{3m}$ . Glucose di- and monophosphates were also obtained by the reaction.

The phosphorylation reaction was examined by varying the molar ratio of  $P_{3m}$  to  $\chi$ -D-glucose (0.5:0.5 mol dm<sup>-3</sup>—— 0.25:2.5 mol dm<sup>-3</sup>), pH (7——14), and

1432 Chemistry Letters, 1987

temperature (room temperature to 70  $^{\circ}$ C). Both  $^{31}$ P-NMR technique and HPLC - FIA (flow injection analysis) system have been applied to determine the structures and the amounts of the products.

Figure 1 shows a representative HPLC profile for the products of  $P_{3m}$  with d-D-glucose after 1 h. In addition to the peaks due to ortho-  $(P_1)$ , pyro-  $(P_2)$ , tri-  $(P_3)$ , and cyclic triphosphate  $(P_{3m})$ , four peaks of unknown compounds (designated as Compounds A, B, C, and D) were observed at 33.2, 16.3, 7.9, and 6.1 min, respectively. By comparing the change in the peak areas with time, it was found that the yield of Compound A increased drastically to reach a maximum value of ca. 47% (as P) after 7 h, and then began to decrease gradually. The amounts of Compounds B, C, and D were small compared with Compound A, though they increased with the progress of the reaction.

The  $^{31}\text{P-NMR}$  spectra of the reaction products shown in Fig. 2 support elucidation of the structures of the unknown compounds. By comparing  $^{1}\text{H-}$  decoupled and  $^{1}\text{H-}$ coupled spectra, we can discriminate phosphorylated glucose compounds from inorganic phosphates ( $P_{1}$ ,  $P_{2}$ ,  $P_{3}$ , and  $P_{3m}$ ).

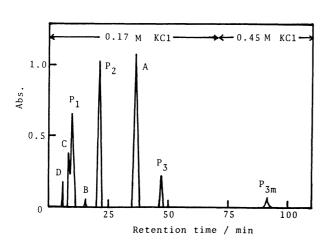


Fig. 1. Elution pattern of the reaction products of &-D-glucose with cyclo-triphosphate.

 $G1u:P_{3m} = 2.5:0.25 \text{ mol dm}^{-3};$ pH 12; at room temperature

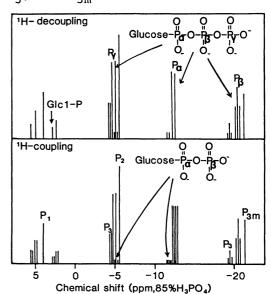


Fig.2. $^{31}$ P-NMR spectra of the reaction products of glucose with  $P_{^3m}$ , Glu:  $P_{^3m}$  = 0.5: 0.5 mol dm $^{-3}$ ; pH 12; at room temperature; after 5h

The peaks at -12.3 ppm (doublet), -21.3 ppm (triplet), and -5.0 ppm (doublet) were assigned to  $P_{\alpha}$ ,  $P_{\beta}$ , and  $P_{\gamma}$  atoms of the triphosphate derivatives of glucose, respectively. The doublet of  $P_{\alpha}$  atom in the <sup>1</sup>H-decoupled spectrum was split into double doublets in the <sup>1</sup>H-coupled spectrum, showing that one proton exists near the  $P_{\alpha}$  atom. It is impossible to define the position of the

Chemistry Letters, 1987

triphosphate group introduced to the glucose molecule by the  $^{31}P\text{-NMR}$  results. Possible compounds are, therefore, glucose 1-, 2-, 3-, and 4-triphosphates. It has been shown that glucose 1-monophosphate was not phosphorylated by  $P_{3m}$ , whereas glucose 6-monophosphate was easily phosphorylated. Also, glucosamine is known to react with  $P_{3m}$  to give a mixture of glucosamine 1-triphosphate and 2-triphosphoramidate. From these additional experimental results, Compound A is expected to be a mixture of glucose 1- and 2-triphosphates.

Two doublets at -11.8 ppm and -5.1 ppm in the <sup>1</sup>H-decoupled spectrum were attributed to P<sub>d</sub> and P<sub>p</sub> atoms of the diphosphate derivative of glucose. The small peak (Compound B) at 12.8 min in the HPLC profile corresponds to glucose diphosphate. By comparing <sup>31</sup>P-NMR spectra of the authentic sample, a singlet 3.5 ppm was assigned to that of glucose 1-monophosphate. Three singlets at 5.3, 5.0, and 3.3 ppm in the <sup>1</sup>H-decoupled spectrum were also attributed to monophosphate derivatives. Compounds C and D, which appeared in the HPLC profile, therefore, correspond to monophosphate derivatives or their mixtures. A further precise study is now in progress in order to clarify the structures of Compounds C and D.

Table 1. Phosphorylation of glucose with cyclo-triphosphate

Reaction	tion conditions		Yield/%		
Molar ratio (Pam: Glu)	рН	Temp /°C	P <sub>3</sub> -derivative	P <sub>1</sub> -derivative	
				(D)	(C)
1:1	12	room temp	16.0	0.2	4.3
	10	room temp	20.0	0.2	1.1
1:5	12	room temp	47.0	1.9	6.7
		70	23.9	0.6	4.7
	10	room temp	32.0	0.7	1.3
1:10	12	room temp	44.3	2.0	6.4
		30	38.7	1.3	7.0
	10	room temp	32.2	0.9	1.3

The yields of the phosphorylated glucose compounds (glucose triphosphate and glucose monophosphate) determined by the HPLC analysis are summarized in Table

1434 Chemistry Letters, 1987

1. The yields of glucose diphosphate were negligibly small, and are not shown in the table.

Optimum conditions for the phosphorylation of glucose with  $P_{3m}$  were the mixing ratio of 5:1—10:1, pH 12, and at room temperature.

## References

- 1) O. T. Quimby and T. J. Flautt, Z. Anorg. Allg. Chem., 296, 220 (1958).
- 2) T. Miyajima, Y. Miyahara, and S. Ohashi, Polyhedron, 1, 425 (1982).
- 3) W. Feldmann, Z. Chem., 1, 26 (1965).
- 4) N. M. Dombrovskii and A. I. Dorosh, Russ. J. Inorg. Chem., 17, 981 (1972).
- 5) W. Feldmann, Z. Chem., 4, 154 (1969).
- 6) J. Rabinowitz, Helv. Chim. Acta, <u>52</u>, 2663 (1969); <u>53</u>, 1350 (1970); <u>54</u>, 1483 (1971).
- 7) J. Rabinowitz, Nature, 224, 795 (1969).
- 8) W. Feldmann, Chem. Ber., 100, 3850 (1967).
- 9) W. Feldmann, Chem. Ber., 99, 3251 (1966).
- 10) A. W. Schwartz, Chem. Commun., 1969, 1393.
- 11) A. W. Schoffstall, Origins Life, <u>7</u>, 399 (1976).
- 12) R. Saffhill, J. Org. Chem., 35, 2881 (1970).
- 13) E. Etaix and L. E. Orgel, J. Carbohydr. Nucleosides, Nucleotides, <u>5</u>, 91 (1978).
- 14) M. Tsuhako, M. Fujimoto, and S. Ohashi, Chem. Lett., 1981, 849.
- 15) M. Tsuhako, M. Fujimoto, S. Ohashi, H. Nariai, and I. Motooka, Bull. Chem. Soc. Jpn., <u>57</u>, 3274 (1984).
- 16) M. Tsuhako, C. Sueyoshi, Y. Baba, T. Miyajima, S. Ohashi, H. Nariai, and I. Motooka, to be published.

(Received April 30, 1987)